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Branched-chain amino acids, hepatocyte growth factor and protein production in the liver

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Abstract

Although the functions associated with differentiation are thought to be suppressed when cells proliferate, recent studies have shown that several mitogens can stimulate functions such as protein production under certain physiological conditions. Hepatocyte growth factor (HGF) is now considered to be a pluripotent factor and has been shown to stimulate the differentiated functions of hepatocytes, as well as their proliferation. The use of HGF for the treatment of liver disease, especially hepatic failure, has been suggested.

Because patients with decompensated liver cirrhosis have decreased plasma concentrations of branched-chain amino acids (BCAAs), many investigations in laboratory animals and patients have been designed to demonstrate the benefits of supplementation of BCAAs on the hepatic metabolism of proteins. However, the mechanisms involved in the specific actions of BCAAs remain to be elucidated. Amino acids are molecules that modulate numerous cellular functions. BCAAs are known to influence gene expression, cellular metabolism, amino acid transport, and protein turnover.

In this paper, we show the potential of BCAAs for stimulating HGF synthesis in the liver and discuss the possibility that BCAAs stimulate protein production by hepatocytes through the induction of HGF.

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Hepatocyte growth factor (HGF) was identified as a potent mitogen for hepatocytes [1–3]. However, recent studies have shown that HGF is pluripotent in its effects [1–7]. Furthermore, the administration of HGF in experimental models of various diseases has been shown to produce favorable effects [2–14]. These studies imply that HGF is a candidate therapeutic agent in many areas of disease. In addition, the up-regulation of the levels and/or activities of HGF in vivo may offer similar benefits.

In patients with liver cirrhosis, plasma concentrations of branched chain amino acids (BCAAs; valine, leucine and isoleucine) are usually reduced, depending on the stage of

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the disease [1,15]. Because a relationship between the occurrence of hepatic encephalopathy and a decrease of BCAAs, as well as an increase of aromatic amino acids, has been shown in the plasma of cirrhotic patients, BCAA-enriched formulas have been administered to such patients to prevent or treat hepatic encephalopathy [16]. Furthermore, the value of BCAA supplements as a nutritional support in cirrhosis, to prevent a decrease in protein production by hepatocytes, has been a topic of study for many years [17-22]. However, previous studies aimed at addressing this question in patients have generated conflicting results. Recently, a double-blind, randomized trial was performed, and some favorable effects on liver function were shown [23]. The potential benefits of supplementation in patients with liver cirrhosis with BCAAs were highlighted but the mechanisms of their action remain to be elucidated. BCAAs are known to be essential for protein

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nutrition. A very simple hypothesis is that supplementation of BCAAs provides the substrates for protein production, resulting in the increased production of proteins by hepatocytes. However, the overall capacity to produce proteins is known to be reduced in the cirrhotic liver, and controversy exists as to whether, and by what mechanism, the supplementation of substrates alone results in an increase in protein synthesis. Other mechanisms also should be considered.

Recent studies have shown that amino acids are molecules that modulate numerous cellular functions. Among them, the BCAAs have been reported to be able to regulate intracellular signaling pathways and gene expression in vitro [1,2,24–36]. The effect of BCAAs on HGF production should be examined as a possible mechanism of stimulation of protein production by hepatocytes, because HGF has the ability to enhance protein production.

1. Pharmacological actions of BCAAs

BCAAs are essential for protein nutrition and are abundant in mammalian proteins; their importance as substrates for protein production is well-established. In addition, leucine and isoleucine are known to be good sources of energy, producing 40 mol of ATP/mole [1,15]. Furthermore, as well as their nutritional aspects, BCAAs have pharmacological and biological activities. They are known to have specific effects in preserving body proteins by increasing the synthesis and decreasing the degradation of protein [1,30,33,36]. Recently, one of the BCAAs, leucine, was reported to activate the phosphorylation of two molecules involved in protein synthesis, p70 S6 kinase and eukaryotic initiation factor 4E binding protein 1 (4E-BP1) [1,25,33-36]. This activation seems to be mediated predominantly by the mammalian target of rapamycin. In theory, all mRNAs can be regulated by such mechanisms. However, some mRNAs are more sensitive to the changes than others, resulting in modulation of gene expression through altered patterns of translation of specific mRNAs [37]. In addition, BCAAs have been shown to have regulatory effects on cellular metabolism as well as amino acid transport. BCAAs can exert pharmacological effects on protein production through many mechanisms.

2. The multifaceted nature of HGF

Although HGF was originally isolated as a stimulator of DNA synthesis by hepatocytes in culture [1-3], it is now recognized as a pluripotent factor acting on various types of cells [1-7]. Epithelial cells initially were identified as the targets of HGF, although non-epithelial cells, such as hematopoietic, lymphoid, neural and muscle cells, also have been shown to respond to HGF. In the liver, HGF stimulates not only the proliferation of hepatocytes but also their differentiated functions, such as protein production, in vitro and in vivo [1-7,11-13]. In addition, HGF administration

suppresses hepatic damage induced by treatment with alphanaphthylisothiocyanate, alcohol, anti-Fas antibody, carbon tetrachloride. dimethylnitrosamine, lipopolysaccharide plus galactosamine, and warm ischemia/reperfusion and/or accelerates recovery from the hepatic injury in experimental models [4-7,11,14]. In such experiments, the contribution of mitogenic, anti-inflamatory, anti-apoptotic and anti-fibrogenetic activities of HGF have been postulated to be beneficial. Similar effects of HGF administration have been observed for other organs. HGF reduces injury to the kidney, lung, brain and heart, and suppresses fibrogenesis in the kidney [4,6,7]. In addition, HGF has been shown to induce angiogenesis and the development of neurons in experimental models [4-10]. HGF has been proposed as a therapeutic agent for liver disease, as well as a variety of disorders of other organs.

3. Stimulation of HGF synthesis

When HGF is administered intravenously or intraperitoneally, its half-life in the blood is quite short; approximately 5 min [3,12]. Thus, repeated injections of HGF would be necessary to maintain elevated levels. Other approaches are required, such as the use of the vectors expressing HGF or stimulation of intrinsic HGF production [14]. Previous reports have shown that many agents induce HGF expression in vitro. Cytokines (tumor necrosis factor α , interleukin 1α , 1β , and 6, and interferon α , β , and γ), growth factors (epidermal growth factor (EGF), fibroblast growth factor, and insulin-like growth factor families), cyclic AMP, phorbol-12-myristate-13-acetate, phorbol 12,13-dibutyrate, prostaglandin E₂, cholera toxin, norepinephrine, low density lipoprotein and injurin have stimulatory effects on HGF production by various cultured cells [38-45]. However, their efficacy in vivo and systemic reactions are still uncertain. The possibility of adverse effects following their administration in vivo should be considered.

4. Stimulation of HGF production by BCAAs

We examined the effect of BCAAs on HGF secretion by hepatic stellate cells (HSCs) because they are a major source of HGF production in the liver [46,47]. Freshly isolated HSCs transform rapidly into myofibroblast-like cells during culture and lose the ability to produce HGF. Therefore, we used an HSC clone (cHSC) that maintains a consistent phenotype during culture [48,49]. The cells were cultured in a medium, deprived of amino acids for 3 h prior to their use in the following experiments. When the medium was changed to Hanks' balanced salt solution (HBSS), which does not contain amino acids, and supplemented with leucine up to 10 mM, the HGF levels in the culture medium increased in a dose-related manner. However, addition of 10 mM valine or isoleucine did not increase the levels of HGF. When cHSC was cultured

in HBSS supplemented with 10 mM leucine, HGF secretion into the medium was enhanced significantly at 18 h. The difference in the HGF levels detected in the medium of the leucine-treated and non-treated cultures were enhanced in a time-dependent manner up to day 3. Under these serumfree culture conditions, proliferation does not occur and the number of cHSC cells remains constant [48,49]. Thus, cHSC produces HGF constantly and leucine enhances this HGF production. The stimulatory effect of leucine on HGF production was observed even when the cells were cultured in Eagle's minimal essential medium, which contains amino acids including BCAAs, supplemented with 0.5 or 10% fetal calf serum, although the effect was less pronounced than that observed in amino acid- and serum-free culture conditions. The mechanisms of the stimulatory effect of leucine on HGF production remain to be elucidated. It is possible that the addition of leucine provides a substrate for HGF production and/or energy to the cells. However, the addition of valine or isoleucine had no significant effect on HGF levels in the medium. Furthermore, a dose-dependent response to leucine addition was observed even when the cells were cultured in medium containing various amino acids and serum, which provide sufficient energy and substrates for cellular functions. Thus, it is less likely that the stimulatory effect of leucine on HGF production depends on the substrate or energy supply provided by leucine. As mentioned above, leucine was reported to activate the phosphorylation of p70 S6 kinase and 4E-BP1 [1,25,33-36]. Leucine may stimulate HGF synthesis by HSCs pharmacologically by enhancing these pathways. In addition, HGF production may be enhanced by leucine in vivo, because it can stimulate HGF production even in the presence of various amino acids and serum in culture medium. BCAA treatment might be a safe and effective way of stimulation of HGF production.

5. A possible mechanism for selecting HGF activities

In clinical practice, only some of the many effects of HGF are desired in the target organs. In patients with liver cirrhosis, the aim of inducing the up-regulation of HGF production would be the stimulation of hepatocyte functions, including protein production. The cytoprotective and mitogenic activities of HGF also might be required in some cases of liver cirrhosis. One possible mechanism for the selection of HGF activities is their regulation downstream of the initial stimulus. To support this hypothesis, effector gene(s) or amplifier(s) could be induced differentially by the stimulation of HGF, depending on the situation or specific requirements. We examined this approach under experimental conditions that show an increase in HGF activity.

In our experiments in vitro, isolated rat hepatocytes were cultured at high and low densities. DNA synthesis increased in a dose-dependent manner in the hepatocytes cultured at low density following the addition of HGF, while the concentrations of albumin and fibrinogen in the medium were not affected. When the hepatocytes were cultured at high density, addition of HGF increased the concentration of albumin or fibrinogen in the medium in a dose-dependent manner but did not affect DNA synthesis. Only in hepatocytes cultured at low density, the activity of transforming growth factor α (TGF α), which is produced by hepatocytes in the liver and is known to be another mitogen for hepatocytes, contributed to the manifestation of the HGF activity, because total cellular TGFa increased in low-density cultured hepatocytes in a dose-related manner following the addition of HGF, and the addition of antisense TGFa mRNA oligonucleotides to the medium inhibited the increase in $TGF\alpha$ expression and DNA synthesis. The increase in DNA synthesis in the presence of HGF was also suppressed by the addition of anti-TGFα rabbit IgG, which does not recognize other ligands for the TGF α /EGF receptor, such as EGF and heparin-binding EGF-like growth factor [50]. Furthermore, we showed that HGF induced transcription factor p53 expression by the hepatocytes, and the suppression of p53 production or action resulted in reduced TGF α expression, followed by a decrease in DNA synthesis by the hepatocytes [51]. Therefore, the mitogenic activity of HGF in cultured hepatocytes seems to be linked to TGF α activity. Recently, similar effects have been reported by Russell and co-workers [52]. They showed that a tyrosine kinase inhibitor of EGF receptor (EGFR), a receptor for the EGF family including TGF α , blocked both basal and ligand-induced tyrosine phosphorylation of the EGFR, but not that of c-met, a receptor for HGF. Pharmacologic inhibition of the EGFR kinase abolished the proliferative actions of HGF. They concluded that the mitogenic effect of HGF might be secondary to increased synthesis or processing of EGFR ligands, such as TGF α . Furthermore, Kojima and coworkers showed that HGF induces branching tubulogenesis in MDCK cells by modulating the activin-follistatin system [53], suggesting the existence of regulatory mechanisms for HGF activity, other than its mitogenic action.

We determined whether similar mechanisms could be observed in vivo. Partial hepatectomy induced an increase in hepatic and circulating levels of HGF, followed by an increase in the production of TGF α and of hepatocyte proliferation in rats [54]. When an anti-TGF α antibody was administered to partially hepatectomized rats, hepatocyte proliferation was reduced, irrespective of any increase in HGF levels [50]. In rats, after sham-operations, hepatic and circulating levels of HGF increased moderately. However, no increases in the TGF α levels and hepatocyte proliferation were observed in these rats [54]. These results suggest that $TGF\alpha$ activity is related to the mitogenic effect of HGF during liver regeneration in rats. In addition, the observations in clinical settings seem to be comparable with these results in rats [55–59]. In patients who received partial hepatectomy, serum HGF levels were increased, followed by an increase in serum levels of TGF α , which reached a maximum later than the serum HGF levels. The maximum levels that were achieved in each case correlated significantly with the resected volume of the liver and the increased volume of the remaining liver. In contrast,



Fig. 1. Schematic potential pathways of BCAA, HGF and protein production, in addition to proliferation of hepatocytes.

TGF α levels did not increase in patients who underwent operations other than partial hepatectomy, while HGF levels did increase moderately. Also, in patients suffering from acute hepatitis, serum HGF levels reached a maximum earlier than the TGF α levels. In these patients, the maximum serum HGF levels correlated positively with the maximum serum TGF α levels, and the TGF α levels correlated with the maximum serum alanine aminotransferase levels observed in each case. Because livers in acute hepatitis may regenerate depending on the degree of liver damage, the TGF α levels should correlate with the degree of liver regeneration in patients with acute hepatitis. Together, these results suggest that serum TGF α levels increase in patients in relation to the extent of liver regeneration after an increase of serum HGF levels.

6. Conclusions and future prospects

BCAAs seem to have significant effects, not only as a nutrient, but also as a drug. BCAAs, especially leucine, stimulate HGF synthesis by hepatic stellate cells. Increased HGF production induced by BCAAs can stimulate protein production by hepatocytes (Fig. 1). Further investigations will be required to elucidate the regulatory mechanisms of HGF induction by BCAAs and the selection of HGF activities, especially in vivo.

References

- Ichihara A. BCA, HGF, and proteasomes. Biochem Biophys Res Commun 1999;266, 647–51.
- [2] Michalopoulos GK, DeFrances MC. Liver regeneration. Science 1997;276:60–6.
- [3] Tsubouchi H. Hepatocyte growth factor for liver disease. Hepatology 1999;30:333–4.
- [4] Zarnegar R, Michalopoulos GK. The many faces of hepatocyte growth factor: from hepatopoiesis to hematopoiesis. J Cell Biol 1995;129:1177–80.
- [5] Fausto N. Liver regeneration. J Hepatol 2000;32:S19-31.

- [6] Matsumoto K, Nakamura T. Emerging multipotent aspects of hepatocyte growth factor. J Biochem 1996;119:591–600.
- [7] Funakoshi H, Nakamura T. Hepatocyte growth factor: from diagnosis to clinical applications. Clin Chim Acta 2003;327:1–23.
- [8] Maina F, Klein R. Hepatocyte growth factor, a versatile signal for developing neurons. Nat Neurosci 1999;2:213–7.
- [9] Bussolino F, Di Renzo MF, Ziche M, et al. Hepatocyte growth factor is a potent angiogenic factor which stimulates endothelial cell motility and growth. J Cell Biol 1992;119:629–41.
- [10] Ware LB, Matthay MA. Keratinocyte and hepatocyte growth factors in the lung: roles in lung development, inflammation, and repair. Am J Physiol 2002;282:L924–40.
- [11] Ishiki Y, Ohnishi H, Muto Y, Matsumoto K, Nakamura T. Direct evidence that hepatocyte growth factor is a hepatotrophic factor for liver regeneration and has a potent antihepatitis effect in vivo. Hepatology 1992;16:1227–35.
- [12] Fujiwara K, Nagoshi S, Ohno A, et al. Stimulation of liver growth by exogenous human hepatocyte growth factor in normal and partially hepatectomized rats. Hepatology 1993;18:1443–9.
- [13] Yamaoka M, Hirata K, Ogata I, et al. Enhancement of albumin production by hepatocyte growth factor in rat hepatocytes: distinction in mode of action from stimulation of DNA synthesis. Liver 1998;18:52–9.
- [14] Ueki T, Kaneda Y, Tsutsui H, et al. Hepatocyte growth factor gene therapy of liver cirrhosis in rats. Nat Med 1999;5:226–30.
- [15] Cooper AJL. Role of the liver in amino acid metabolism. In: Zakim T, Boyer TD, editors. Hepatology: a text book of liver disease. Philadelphia: W.B. Saunders company; 1996. p. 563–601.
- [16] Fabbri A, Magrini N, Bianchi G, Zoli M, Marchesini G. Overview of randomized clinical trials of oral branched-chain amino acid treatment in chronic hepatic encephalopathy. J Parenter Enteral Nutr 1996;20:159–64.
- [17] Watanabe A, Wakabayashi H, Kuwabara Y. Nutrient-induced thermogenesis and protein-sparing effect by rapid infusion of a branched chain-enriched amino acid solution to cirrhotic patients. J Med 1996;27:176–82.
- [18] Plauth M, Merli M, Kondrup J, Weimann A, Ferenci P, Muller MJ, ESPEN Consensus Group. ESPEN guidelines for nutrition in liver disease and transplantation. Clin Nutr 1997;16:43–55.
- [19] Moriwaki H, Tajika M, Miwa Y, et al. Nutritional pharmacotherapy of chronic liver disease: from support of liver failure to prevention of liver cancer. J Gastroenterol 2000;35:13–7.
- [20] Marchesini G, Bianchi G, Rossi B, Brizi M, Melchionda N. Nutritional treatment with branched-chain amino acids in advanced liver cirrhosis. J Gastroenterol 2000;35:7–12.
- [21] Yamauchi M, Takeda K, Sakamoto K, Ohata M, Toda G. Effect of oral branched chain amino acid supplementation in the late evening on the nutritional state of patients with liver cirrhosis. Hepatol Res 2001;21:199–204.
- [22] Nishiguchi S, Shiomi S, Kawamura E, et al. Evaluation of ammonia metabolism in the skeletal muscles of patients with cirrhosis using N-13 ammonia PET. Ann Nucl Med 2003;17:417–9.
- [23] Marchesini G, Bianchi G, Merli M, et al. Nutritional supplementation with branched-chain amino acids in advanced cirrhosis: a doubleblind, randomized trial. Gastroenterology 2003;124:1792–801.
- [24] Hutson SM, Harris RA. Symposium—leucine as a nutritional signal: introduction. J Nutr 2001;131:839–40.
- [25] Gautsch TA, Anthony JC, Kimball SR, Paul GL, Layman DK, Jefferson LS. Availability of eIF4E regulates skeletal muscle protein synthesis during recovery from exercise. Am J Physiol 1998;274:C406–14.
- [26] Yoshizawa F, Kimball SR, Jefferson LS. Modulation of translation initiation in rat skeletal muscle and liver in response to food intake. Biochem Biophys Res Commun 1997;240:825–31.
- [27] Harris RA, Kobayashi R, Murakami T, Shimomura Y. Regulation of branched-chain α-keto acid dehydrogenase kinase expression in rat liver. J Nutr 2001;131:841S–5S.

- [28] Flakoll PJ, Wentzel LS, Rice DE, Hill JO, Abumrad NN. Shortterm regulation of insulin-mediated glucose utilization in four-day fasted human volunteers: role of amino acid availability. Diabetologia 1992;35:357–66.
- [29] McDowell HE, Christie GR, Stenhouse G, Hundal HS. Leucine activates system A amino acid transport in L6 rat skeletal muscle cells. Am J Physiol 1995;269:C1287–94.
- [30] May ME, Buse MG. Effects of branched-chain amino acids on protein turnover. Diabetes Metab Rev 1989;5:227–45.
- [31] Louard RJ, Barrett EJ, Gelfand RA. Overnight branched-chain amino acid infusion causes sustained suppression of muscle proteolysis. Metabolism 1995;44:424–9.
- [32] Laine RO, Hutson RG, Kilberg MS. Eukaryotic gene expression: metabolite control by amino acids. Prog Nucleic Acid Res Mol Biol 1996;53:219–48.
- [33] Peyrollier K, Hajduch E, Blair AS, Hyde R, Hundal HS. L-Leucine availability regulates phosphatidylinositol 3-kinase, p70 S6 kinase and glycogen synthase kinase-3 activity in L6 muscle cells: evidence for the involvement of the mammalian target of rapamycin (mTOR) pathway in the L-leucine-induced up-regulation of System A amino acid transport. Biochem J 2000;350:361–8.
- [34] Shigemitsu K, Tsujishita Y, Miyake H, et al. Structural requirement of leucine for activation of p70 S6 kinase. FEBS Lett 1999;447:303–6.
- [35] Xu G, Kwon G, Marshall CA, Lin TA, Lawrence Jr JC, McDaniel ML. Branched-chain amino acids are essential in the regulation of PHAS-I and p70 S6 kinase by pancreatic beta-cells – a possible role in protein translation and mitogenic signaling. J Biol Chem 1998;273:28178–84.
- [36] Anthony JC, Lang CH, Crozier SJ, et al. Contribution of insulin to the translational control of protein synthesis in skeletal muscle by leucine. Am J Physiol Endocrinol Metab 2002;282:E1092–101.
- [37] Kimball SR, Jefferson LS. Regulation of global and specific mRNA translation by oral administration of branched-chain amino acids. Biochem Biophys Res Commun 2004;313:423–7.
- [38] Gohda E, Kataoka H, Tsubouchi H, Daikilara Y, Yamamoto I. Phorbol ester-induced secretion of human hepatocyte growth factor by human skin fibroblasts and its inhibition by dexamethasone. FEBS Lett 1992;301:107–10.
- [39] Tamura M, Arakaki N, Tsubouchi H, Takada H, Daikuhara Y. Enhancement of human hepatocyte growth factor production by interleukin-1 alpha and -1 beta and tumor necrosis factor-alpha by fibroblasts in culture. J Biol Chem 1993;268:8140–5.
- [40] Inaba M, Koyama H, Hino M, et al. Regulation of release of hepatocyte growth factor from human promyelocytic leukemia cells, HL-60, by 1,25-dihydroxyvitamin D3, 12-O-tetradecanoylphorbol 13-acetate, and dibutyryl cyclic adenosine monophosphate. Blood 1993;82:53–9.
- [41] Liu Y, Michalopoulos GK, Zarnegar R. Structural and functional characterization of the mouse hepatocyte growth factor gene promoter. J Biol Chem 1994;269:4152–60.
- [42] Gohda E, Matsunaga T, Kataoka H, Takebe T, Yamamoto I. Induction of hepatocyte growth factor in human skin fibroblasts by epidermal growth factor, platelet-derived growth factor and fibroblast growth factor. Cytokine 1994;6:633–40.
- [43] Broten J, Michalopoulos G, Petersen B, Cruise J. Adrenergic stimulation of hepatocyte growth factor expression. Biochem Biophys Res Commun 1999;262:76–9.

- [44] Matsumoto K, Tajima H, Hamanoue M, Kohno S, Kinoshita T, Nakamura T. Identification and characterization of 'injurin', an inducer of expression of the gene for hepatocyte growth factor. Proc Natl Acad Sci USA 1992;89:3800–4.
- [45] Haug C, Schmid-Kotsas A, Zorn U, et al. Hepatocyte growth factor is upregulated by low-density lipoproteins and inhibits endothelin-1 release. Am J Physiol 2000;279:H2865–71.
- [46] Tomiya T, Inoue Y, Yanase M, et al. Leucine stimulates the secretion of hepatocyte growth factor by hepatic stellate cells. Biochem Biophys Res Commun 2002;297:1108–11.
- [47] Tomiya T, Omata M, Fujiwara K. Significance of branched chain amino acids as possible stimulators of hepatocyte growth factor. Biochem Biophys Res Commun 2004;313(4):11–6.
- [48] Greenwel P, Schwartz M, Rosas M, Peyrol S, Grimaud JA, Rojkind M. Characterization of fat-storing cell lines derived from normal and CCl4-cirrhotic livers. Differences in the production of interleukin-6. Lab Invest 1991;65:644–53.
- [49] Greenwel P, Rubin J, Schwartz M, Hertzberg EL, Rojkind M. Liver fat-storing cell clones obtained from a CCl4-cirrhotic rat are heterogeneous with regard to proliferation, expression of extracellular matrix components, interleukin-6, and connexin 43. Lab Invest 1993;69:210–6.
- [50] Tomiya T, Ogata I, Yamaoka M, Yanase M, Inoue Y, Fujiwara K. The mitogenic activity of hepatocyte growth factor on rat hepatocytes is dependent upon endogenous transforming growth factor α. Am J Pathol 2000;157:1693–701.
- [51] Inoue Y, Tomiya T, Yanase M, et al. P53 may positively regulate hepatocyte proliferation in rats. Hepatology 2002;36:336–44.
- [52] Scheving LA, Stevenson MC, Taylormoore JM, Traxler P, Russell WE. Integral role of the EGF receptor in HGF-mediated hepatocyte proliferation. Biochem Biophys Res Commun 2002;290:197– 203.
- [53] Maeshima A, Zhang YQ, Furukawa M, Naruse T, Kojima I. Hepatocyte growth factor induces branching tubulogenesis in MDCK cells by modulating the activin-follistatin system. Kidney Int 2000;58:1511–22.
- [54] Tomiya T, Ogata I, Fujiwara K. Transforming growth factor alpha levels in liver and blood correlate better than hepatocyte growth factor with hepatocyte proliferation during liver regeneration. Am J Pathol 1998;153:955–61.
- [55] Tomiya T, Nagoshi S, Fujiwara K. Significance of serum human hepatocyte growth factor levels in patients with hepatic failure. Hepatology 1992;15:1–4.
- [56] Tomiya T, Tani M, Yamada S, Hayashi S, Umeda N, Fujiwara K. Serum hepatocyte growth factor levels in hepatectomized and non-hepatectomized surgical patients. Gastroenterology 1992;103:1621–4.
- [57] Tomiya T, Fujiwara K. Serum levels of transforming growth factor α in patients after partial hepatectomy as determined by an enzyme linked immunosorbent assay. Hepatology 1993;18:304–8.
- [58] Tomiya T, Fujiwara K. Liver regeneration in fulminant hepatitis as evaluated by serum transforming growth factor α levels. Hepatology 1996;23:253–7.
- [59] Tomiya T, Hayashi S, Yanase M, et al. Serum transforming growth factor α level can be a parameter for liver regeneration after partial hepatectomy in patients with liver cancer. Semin Oncol 1997;24:14–7.